

Evolution of pollination niches and floral divergence in the generalist plant *Erysimum mediohispanicum*

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- **Background and Aims** How generalist plants diverge in response to pollinator selection without becoming specialized is still unknown. This study explores this question, focusing on the evolution of the pollination system in the pollination generalist *Erysimum mediohispanicum* (Brassicaceae).
- **Methods** Pollinator assemblages were surveyed from 2001 to 2010 in 48 geo-referenced populations covering the entire geographic distribution of *E. mediohispanicum*. Bipartite modularity, a complex network tool, was used to find the pollination niche of each population. Evolution of the pollination niches and the correlated evolution of floral traits and pollination niches were explored using within-species comparative analyses.
- **Key Results** Despite being generalists, the *E. mediohispanicum* populations studied can be classified into five pollination niches. The boundaries between niches were not sharp, the niches differing among them in the relative frequencies of the floral visitor functional groups. The absence of spatial autocorrelation and phylogenetic signal indicates that the niches were distributed in a phylogeographic mosaic. The ancestral *E. mediohispanicum* populations presumably belonged to the niche defined by a high number of beetle and ant visits. A correlated evolution was found between pollination niches and some floral traits, suggesting the existence of generalist pollination ecotypes.
- **Conclusions** It is conjectured that the geographic variation in pollination niches has contributed to the observed floral divergence in *E. mediohispanicum*. The process mediating this floral divergence presumably has been adaptive wandering, but the adaptation to the local pollinator faunas has been not universal. The outcome is a landscape where a few populations locally adapted to their pollination environment (generalist pollination ecotypes) coexist with many populations where this local adaptation has failed and where the plant phenotype is not primarily shaped by pollinators.

Key words: Adaptive wandering, generalist pollination ecotype, pollination niches, floral divergence, *Erysimum mediohispanicum*, Brassicaceae, speciation.

INTRODUCTION

The evolution of floral diversity has been considered mostly as the adaptive outcome of specialized interactions between flowers and their major pollinators (Darwin, 1862; Harder and Johnson, 2009; Willmer, 2011). As a consequence of adaptation to the same functional group of pollinators, plants exhibit a similar suite of covarying floral traits that heighten the attraction and pollen transfer of that specific group of pollinators, the so-called pollination syndrome (Faegri and van der Pijl, 1979; Fenster *et al.*, 2004; Willmer, 2011). Within this theoretical framework, the origin of floral diversity is mediated by the divergent evolution that occurs when a plant species pollinated by one group of pollinators shifts to a different group of pollinators, i.e. when there is a transition between pollination syndromes (Castellanos *et al.*, 2003; Thomson and Wilson, 2008; Harder and Johnson, 2009). In contrast to specialist plants, generalist plants are simultaneously visited by a diverse assemblage of floral visitors with different morphologies, foraging behaviour, preference patterns and per-visit effectiveness (Gómez and Zamora, 1999). As different types of floral visitors show distinct preferences for floral traits and contrasting morphological fitting with floral traits, thereby imposing conflicting selections, the

floral traits of most generalist plants are either not adapted to any particular pollinator or reflect adaptation to multiple pollinator groups (Herrera, 1996; Wilson *et al.*, 2004; Sahli and Conner, 2011). For this reason, generalist plants do not belong to any pollinator syndrome, and most pollination biologists question the role of pollinators as evolutionary drivers in generalist systems (Waser, 2001; Kay and Sargent, 2009).

Pollination systems may be considered as ecological niches (Armbruster *et al.*, 1994; Johnson, 2010; Pauw, 2013). Pollination niches occur in both generalist and specialist plant species, the only difference between them being the breadth of the niches: narrow in specialist plants but wide in generalist plants. Whereas the boundaries between pollination niches in specialist plants are sharp, generalist plant species differ from each other in the relative frequency of different types of pollinators visiting their flowers (Grant and Grant, 1965; Armbruster, 1988, 1993; Dilley *et al.*, 2000; Thomson and Wilson, 2008). Differences among generalist plants in pollination niches are quantitative rather than qualitative. Although pollination niches have been studied mostly to investigate the structure of plant communities and the possibility of plant coexistence under interspecific competition (Parrish and Bazzaz, 1978,

1979; Armbruster *et al.*, 1994; Abe, 2006; Ollerton *et al.*, 2007; Geib and Galen, 2012), they can also be useful to understand floral divergence. In fact, because of the tight equivalence between pollination niche (the array of animals visiting the flowers) and syndrome (the suite of floral traits displayed by the plant to attract those animals) in specialist plants, pollinator-driven floral diversification occurs in these plants when there is a transition between pollination niches. A similar process might also occur in generalist plants, where floral divergence could be driven by shifts between different generalist pollination niches. This possibility for pollinator-mediated divergence in floral morphology, however, has scarcely been explored in generalist clades (Dilley *et al.*, 2000; Sargent and Otto, 2006; Medel *et al.*, 2007; Cooley *et al.*, 2008; Smith *et al.*, 2008).

Scaling down from species to populations, pollinator identity and abundance also vary between conspecific populations in most generalist plants (Aigner, 2005; Moeller, 2005; Price *et al.*, 2005; and references therein). This quantitative variation in pollinator fauna creates concomitant spatial variation in the selection undergone by plants (Galen and Cuba, 2001; Thomson and Wilson, 2008) and could ultimately cause phenotypic divergence, mostly in geographically isolated populations where gene flow is disrupted (Waser, 2001; Kay and Sargent, 2009). A potential consequence of this between-population variation in pollination niche is the evolution of pollination ecotypes, floral variants adapted to the local pollinator fauna (Grant and Grant, 1965; Armbruster, 1993; Johnson, 2006, 2010; Pérez-Barrales *et al.*, 2007; Andersson *et al.*, 2009; Armbruster and Muchhala, 2009). Pollination ecotypes may be interpreted as a path to the specialization in a sub-set of functionally similar pollinators (Fenster *et al.*, 2004; Johnson, 2010; van der Niet *et al.*, 2014). Whether generalist plants diverge in response to changes in pollination niches and develop different pollination ecotypes without becoming specialized is still an open question. Thomson and Wilson (2008; see also Dilley *et al.*, 2000) have envisaged a potential process, called adaptive wandering, to explain the pollinator-mediated phenotypic divergence of generalist flowers. They posit a geographical divergence in plant phenotype as a consequence of local adaptation to slightly different selection regimes, as imposed by local pollinator communities. Plants adapt to new local pollinators without excluding other pollinators. This would eventually result in new plant species adapting to slightly different but still diverse pollinator fauna, i.e. adaptive wandering (Thomson and Wilson, 2008). Since adaptive wandering does not involve a shift towards a new functional group of pollinators, floral traits should vary in generalist clades not in absolute terms but gradually and continuously (Armbruster, 1993; Armbruster and Muchhala, 2009).

The main aim of this study is to investigate the role played by pollinators in the phenotypic divergence of a generalist plant species, *Erysimum mediohispanicum* (Brassicaceae). The main hypotheses we will check in this study are as follows. (1) Although pollinators vary geographically, they are not distributed randomly across plant populations; consequently, there exists intraspecific, between-population variation in pollination niches. (2) These pollination niches are generalist, depending exclusively on the spatial variation in the relative abundance of some pollinators rather than in the specialization of some plant populations to sub-sets of pollinators. (3) There is geographic and phylogeographic structure in pollination niche variation,

with nearby and genetically similar populations belonging to the same pollination niches. (4) Populations belonging to the same pollination niche display similar floral traits and can be considered generalist pollination ecotypes. We have tested these hypotheses by means as follows. Using tools from the network theory, we first identify the pollination niches of this plant species, based on the relative abundance of different types of flower visitors. We then determine the spatial, between-population variation of these pollination niches throughout the entire geographic distribution of the plant. We subsequently infer how these pollination niches have evolved over the course of the plant's evolutionary history. Finally, we quantify the phylogeographic association between the pollination niches and the floral phenotype of the plants using within-species comparative analyses. Collectively, this information will provide a framework for future investigations on pollinator-mediated phenotypic evolution and divergence of generalist plants.

MATERIALS AND METHODS

Plant natural history

Erysimum mediohispanicum (Brassicaceae) is a mostly biennial, monocarpic herb endemic to the Iberian Peninsula, where it occupies two separate regions, one in the north and the other in the south-east of Spain (Nieto-Feliner, 1993). Individual plants grow for 2–3 years as vegetative rosettes, and then die after producing 1–8 reproductive stalks bearing up to several hundred hermaphroditic, bright-yellow flowers containing 30–40 ovules (Gómez *et al.*, 2009a). *Erysimum mediohispanicum* is partially self-compatible but requires pollen vectors for full seed set. The pollination system of this species is extremely generalist (Gómez *et al.*, 2007), its flowers being visited by several hundred insect species over its entire distribution area (Gómez *et al.*, 2007, 2009a). Despite this generalization, pollinators act as effective selective pressures on several floral traits (Gómez *et al.*, 2008a, 2009a). Previous studies have demonstrated that spatial variation in pollinators causes concomitant geographical variation in the selection acting on *E. mediohispanicum* floral traits (Gómez *et al.*, 2008a, 2009a), which can result in patterns of local adaptation to pollinator fauna (Gómez *et al.*, 2009b).

Sampling design

During the years 2001–2010, we studied 48 geo-referenced populations spanning the entire distribution of *E. mediohispanicum*. To do so 14 populations were sampled in two areas from the northern region, one in the Iberian Plateau including localities in Guadalajara, Soria and Zaragoza provinces, and the other in the Pre-Pyrenees, including localities in the Sierra del Montsec (Lleida province) and Sierra de Guara (Huesca; Supplementary Data Table S1). We also sampled 34 populations in two areas from the southern region: the Prebaetic System, including localities in Sierras de Cazorla, Segura y Las Villas (Jaén), Sierra de la Guillimona (Granada) and Sierra de Alcaraz (Albacete); and the Penibaetic System, including localities in Sierra Nevada (Granada and Almería), Sierra de Cogollos (Granada), Sierra de Lújar (Granada), Sierra de Gádor (Almería) and Sierra de Espuña (Murcia) (Fig. 1; Supplementary Data Table S1). The genetic relationship of these populations was inferred from the

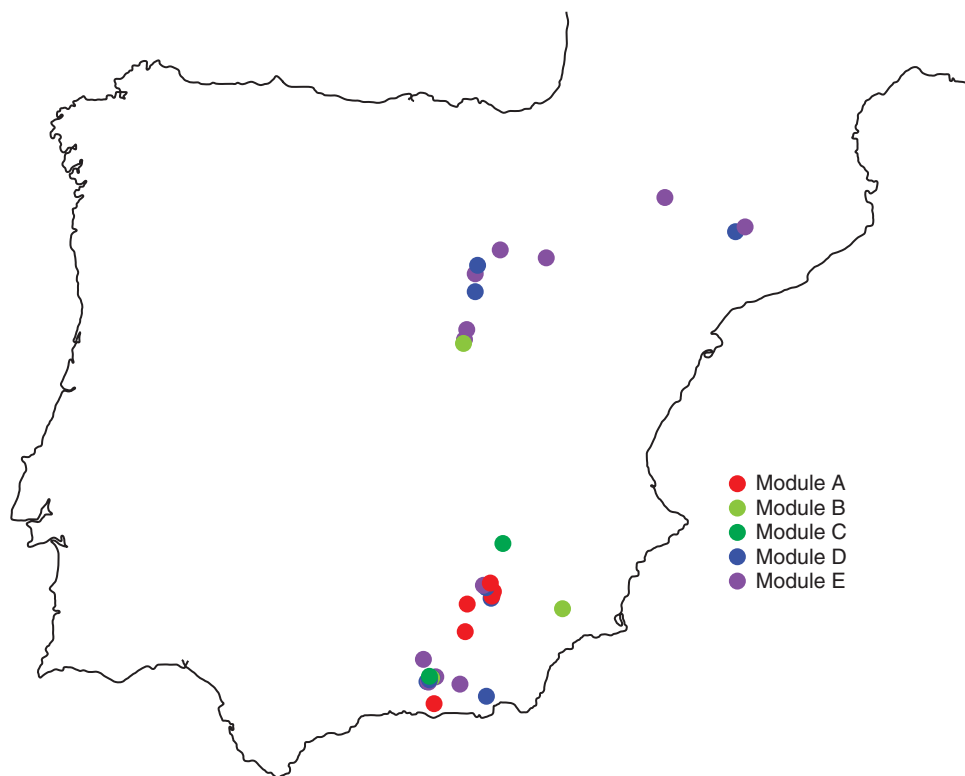


FIG. 1. Geographic distribution of the 48 populations of *Erysimum mediohispanicum* studied in the Iberian Peninsula. Each colour represents a different module (see key).

trnL-trnF intergenic spacer (IGS) chloroplast DNA (cpDNA) region using a new method developed by Muñoz-Pajares (2013) that combines the information from indels and substitutions (see Supplementary Data Methods for a full description of the methodology).

Plant phenotypic traits

The phenotype of at least 30 individual plants per population and year (4316 plants in total; Supplementary Data Table S1) was characterized, quantifying the following phenotypic traits: (1) stalk height, i.e. the height in centimetres of the tallest flowering stalk; (2) flower number, i.e. the total number of flowers and floral buds produced by each plant; (3) corolla diameter, i.e. the distance in millimetres between the apical edges of two opposite petals; (4) corolla tube length, i.e. the distance in millimetres between the corolla tube aperture and the base of the sepals; and (5) corolla shape, i.e. a multidimensional trait estimated by geometric morphometric tools using a landmark-based methodology (Zelditch et al., 2004). For each individual, we selected one flower at anthesis and took a digital photo of the front view and planar position. We defined 32 co-planar landmarks covering the corolla shape and using midrib, primary and secondary veins, and petal edges and connections (see Gómez and Perfectti, 2010 for a full description of the methodology applied to *E. mediohispanicum*). Using the two-dimensional co-ordinates of landmarks, the generalized orthogonal least-squares Procrustes average was computed using the Generalized Procrustes Analysis (GPA) superposition method. To visualize

shape differences, we computed the relative warps (RWs), which are the principal components of the covariance matrix of the partial warp scores and uniform components (Adams et al., 2004). Although our set of 32 landmarks generated 60 RWs, we used the first four RWs in all subsequent analyses because they were the only ones that separately explained >5% of the total variation in shape (Gómez and Perfectti, 2010). RW1 is associated with changes in petal parallelism, RW2 with dorsoventral asymmetry, RW3 with left and right petal asymmetry and RW4 with changes in corolla roundness (Supplementary Data Fig. S1).

Pollinator surveys

Pollinator counts were conducted for at least 2 years in most populations (Supplementary Data Table S1). We visited each population during bloom peak and recorded the insects visiting the flowers, recording only those insects that made contact with the anthers or stigma at least during part of their visit to the flowers (i.e. they could act as pollinators). Previous studies in *E. mediohispanicum* (Gómez et al., 2009a) indicate that 130–150 insects per population provided an accurate estimate of the local pollinator assemblage. For this reason, each population was visited several times to ensure this sampling effort (Supplementary Data Table S1). Unfortunately, it was impossible to reach this amount of floral visitors in a few populations where insects were especially scarce. However, we kept these populations in our study because it was observed that their removal did not change our main outcomes. Pollinators were

identified in the field, and some specimens were captured for further identification in the laboratory. Some rare pollinators could not be captured and we could thus identify them only to genus or family (see Gómez et al., 2007, for details).

We grouped the insects visiting *E. mediohispanicum* flowers into functional groups (Fenster et al., 2004). A ‘functional group’ is defined here as those insects that interact with the flowers in a similar manner. Basically, we used criteria of similarity in body length, proboscis length, foraging behaviour and feeding habits. Thus, taxonomically related species were sometimes placed in different functional groups. Sixteen functional groups were established. (1) Long-tongued large bees: mostly nectar-collecting females ≥ 10 mm in body length belonging to the families Anthophoridae (mostly *Anthophora* spp.) and Apidae (*Apis mellifera* and several *Bombus* spp.). (2) Short-tongued large bees: mostly pollen- and nectar-collecting females > 10 mm belonging primarily to the families Halictidae (*Lasioglossum* spp., *Halictus* spp.), Megachilidae (*Osmia* spp.), Colletidae (*Colletes* spp.) and Andrenidae (*Andrena* spp.). (3) Short-tongued medium-sized bees: mostly pollen- and nectar-collecting females between 5 and 10 mm also belonging to the families Halictidae (*Lasioglossum* spp., *Halictus* spp.) and Andrenidae (*Andrena* spp.). (4) Short-tongued small bees: mostly pollen- and nectar-collecting females < 5 mm. Although they were pollinators, they could act as nectar thieves and belonged primarily to the families Halictidae (*Lasioglossum* spp.), Colletidae (*Hyleaus* spp.), Andrenidae (*Andrena* spp.), Apidae Xylocopinae (*Ceratina* spp.) and Apidae Nomidinae (*Nomada* spp.). (5) Ants: both orthodox pollinators and nectar thieves belonging mostly to the genera *Formica*, *Camponotus*, *Proformica*, *Plagiolepis* and *Leptothorax*. (6) Large wasps: large aculeate wasps, parasitic wasps and kleptoparasitic bees collecting only nectar (mostly *Polistes* spp.). (7) Small wasps: small parasitic wasps belonging to Chalcidoidea and Ichneumonoidea, collecting only nectar, and acting both as pollinators and nectar thieves. (8) Beeflies: long-tongued nectar-collecting flies belonging to the families Bombyliidae (mostly *Bombylius* spp.) and Nemestrinidae. (9) Hoverflies: nectar- and pollen-collecting Syrphidae and short-tongued Bombyliidae. (10) Large flies: nectar-collecting flies > 5 mm, mostly belonging to the families Muscidae, Calliphoridae, Tabanidae, Scatophagidae and Anthomyiidae. (11) Small flies: nectar-collecting flies < 5 mm mostly belonging to families Muscidae, Anthomyiidae, Micetophyllidae, Empididae, Bibionidae, Drosophilidae and Stratiomyidae (these flies, although pollinating the flowers, may also act also as nectar thieves). (12) Beetles: including species collecting nectar and/or pollen mostly belonging to the families Melyridae (Malachidae and Dasytidae), Cleridae, Oedemeridae, Nitidulidae, Elateridae, Bruchidae, Buprestidae, Phalacridae and Chrysomelidae. (13) Butterflies: mostly Rhopalocera belonging to the families Nymphalidae and Pieridae plus some diurnal moths belonging to the family Sphingidae, all nectar collectors. (14) Moths: small nectar-collecting Lepidoptera mostly belonging to the families Adelidae and Incurvariidae. (15) Bugs: nectar-collecting Hemiptera belonging mostly to the family Lygaeidae and Pentatomidae (outstanding *Eyridema* spp.) (these insects also act as sap suckers). (16) Others: some species of grasshoppers, snakeflies, thrips and earwigs that visit the flowers to collect both pollen and nectar.

Determination of pollination niches

The occurrence of different pollination niches in our study populations was determined using bipartite modularity, a complex network metric. An ecological network is modular when it can be sub-divided into groups (= modules) sharing most of their interactions with members in the same group and a few interactions with members from other groups (Guimerà et al., 2007). Network modularity is a measure of the extent to which species have more links within their modules than would be expected if the linkage were random.

A weighted bipartite network was constructed that included all the populations and the frequency of visits of each pollinator functional group per population. In this network, we pooled the data from the whole study period (2001–2010) and did not consider the time difference involved in sampling across different populations. We subsequently determined the modularity level in this weighted bipartite network using the QuaBiMod algorithm (Dormann and Strauss, 2013). This method used a recurrent Markov chain Monte Carlo (MCMC) algorithm to find the best division of populations into modules. A total of 10^6 MCMC steps were used with a tolerance level of 10^{-10} . As random networks may also have strong modularity, we explored whether our network was significantly more modular than random networks by running the same algorithm in 100 random networks, with the same linkage density as the empirical one (Guimerà and Amaral, 2005). Modularity significance was tested by comparing the empirical vs. the random modularity indices (Guimerà and Amaral, 2005). After testing the modularity of our network, we determined the number of modules using the approach proposed by Newman (2004). We subsequently identified the floral visitor functional groups, defining each module and the populations that were ascribed to each module. To check for uncertainty in module identity due to method accuracy, we repeated the analysis 50 times and recorded the distribution of populations across modules each time. Thus, for each population, we weighed the probability of belonging to each module. Furthermore, we tested for temporal consistency in module identity by performing a separate modularity analysis for every year in which we had sampled > 10 populations (2007, 2008, 2009 and 2010). All modularity analyses were performed using R package bipartite 1.18 (Dormann and Gruber, 2012).

Spatial structure of pollination niches

We tested whether the pollinator fauna of *E. mediohispanicum* was spatially structured by testing variation between regions (south vs. north) and areas (Prebaetic System, Penibaetic System, Iberian Plateau and Pre-Pyrenees) in module identity by means of a nested generalized linear mixed model (glmm). The dependent variable (module identity) was fitted to a multinomial, and area was considered a random factor nested onto region. This analysis was performed using the R package lme4 0.99-0 (Bates, 2005).

Comparative analysis across *Erysimum mediohispanicum* populations

In this study, we test whether pollination niches depend on the genetic relatedness of the populations (phylogenetic signal),

inferring the characteristics of the most likely pollination niche in *E. mediohispanicum* ancestral populations, and exploring the correlated evolution between pollination niches and floral traits (pollination ecotypes). Population traits are not statistically independent because they include a component inherited from ancestral populations plus a component owing to gene flow with other populations (Stone *et al.*, 2011). Across-population analyses need to cope with these sources of non-independence. However, as opposed to cross-species comparative analyses, the within-species comparative analysis is not yet fully developed (Stone *et al.*, 2011; Mardulyn, 2012). We tried to solve this problem by using the between-population genetic distance matrix established with the trnL-trnF IGS cpDNA region to control for across-population non-independence when possible.

Phylogenetic signal. Phylogenetic signal in pollinators was tested by means of the Mantel test (Hardy and Pavoine, 2012; Pavoine and Ricotta, 2013). We correlated the genetic distance matrix with a matrix containing the between-population pollinator distances determined using the Bray–Curtis dissimilarity index (Pavoine and Ricotta, 2013). In addition, we included the geographic distance matrix to control for spatial location of the populations.

Ancestral reconstruction of pollination niches. Unfortunately, methods to estimate ancestral states are not available for network topologies and this requires the use of phylogenetic trees (Nunn, 2011). For this reason, the matrix of between-population relatedness was converted into a phylogenetic tree (see Honnay *et al.*, 2009 for a similar approach with *Erysimum cheiranthoides*). Nevertheless, we exercised caution when interpreting the outcomes from this analysis, since we were assuming no between-population gene flow.

The genetic relationships between *E. mediohispanicum* populations were represented by means of the Neighbor–Joining (NJ) algorithm. Ancestral states were estimated using an updated version of the ‘ancThresh’ command implemented in the R package phytool 0.2-14 (Revell, 2012). This function uses Bayesian MCMC to estimate ancestral states and the threshold model to establish the thresholds for a discrete character to change between states (Felsenstein, 2012). Under the threshold model, the evolving discrete trait is considered to have a continuous, underlying liability (Felsenstein, 2012). When the liability exceeds a threshold value, the discretely valued state of the observable character trait changes. Moreover, this method makes it possible to find the ancestral states, either by considering the state of the tips as a fixed value or by considering uncertainty in character state by assigning to each tip a prior probability of belonging to any of the states conforming the character (Revell, 2012). We followed both of these approaches. Thus, we first reconstructed the ancestral module identity by considering the module identity of the *E. mediohispanicum* populations as a fixed value. We then reconstructed the ancestral state in module identity by considering the uncertainty in ascribing the populations to different modules. To do this, we reran the ancestral reconstruction analysis but included the possibility that some populations belonged to different modules (see above). Finally, we also reconstructed ancestral state in module identity, taking into account the uncertainty in ascribing the populations to different modules as a consequence of temporal changes in pollinator fauna. Again, we reran the ancestral reconstruction analysis but included the possibility of some populations of changing

module identity between years (see above). In all cases, we ran four chains of 10^6 generations each. The posterior probabilities for each character state at each internal node were determined with the ‘ace’ command in R packages ape 3.0-6 (Paradis *et al.*, 2004) and phytool 0.2-14 (Revell, 2012).

Correlated evolution of pollination niches and phenotypic traits. The correlated evolution of pollination niche and plant phenotype was explored using MCMCglmm (Hadfield and Nakagawa, 2010; Stone *et al.*, 2011). MCMCglmm can incorporate the between-population relatedness due to migration and population history as a correlated random effect (Hadfield, 2013). In the models, we included as a random variance structure both the genetic distance matrix and the geographic distance matrix across populations. The dependent variable, module identity, was considered a non-ordered multinomial variable. We used 10^5 MCMC steps with a burn-in of 50 000 and a thinning interval of 10. All the analyses were performed using the R package MCMCglmm 2.17 (Hadfield, 2013).

RESULTS

Determining pollination niches

A total of 6442 flower visits from 545 insect morphospecies belonging to 99 families and seven orders were recorded. The average number of insect species (\pm s.e.) recorded per population was 28.6 ± 1.46 . All the populations studied were generalists. The most frequent floral visitors belonged to the orders Coleoptera (40 % of the visits), Hymenoptera (28 %), Diptera (17 %) and Lepidoptera (9 %). The rest of the insect orders accounted for <5 % of the visits. However, the most frequent functional groups were beetles, long-tongued large bees, short-tongued large bees, short-tongued medium-sized bees, short-tongued small bees, beeflies and butterflies (Supplementary Data Table S2).

The network between *E. mediohispanicum* populations and the pollinator functional groups was significantly modular [empirical modularity \pm s.e. = 0.311 ± 0.001 ; random modularity \pm s.e. (95 % confidence interval) = 0.0301 ± 0.0003 (0.0300–0.0302); $n = 100$ replicates]. This analysis detected five modules, depending on the frequency of the different groups of floral visitors per population (Fig. 2): module A, defined by medium-sized bees and, to a lesser extent, small flies and moths; module B, defined by long-tongued large bees and, to a lesser extent, large wasps, hoverflies and large flies; module C, defined by butterflies and, to a lesser extent, small wasps; module D, defined mostly by beeflies, short-tongued small and large bees, and bugs; and module E, defined by beetles and ants. The average number of functional groups found per population was 8.7 ± 0.5 . Overall, the number of populations belonging to each module was heterogeneous. Whereas modules A, B and C were composed of five, seven and three populations, respectively, module D was composed of 13 populations, and module E was composed of 20 populations (Table 1, Fig. 2).

The module identity of the populations was very consistent. Thus, only four populations (Em10, Em27, Em42 and Em52) changed considerably between modules when the analysis was repeated 50 times, whereas the remaining 44 populations were consistently ascribed to a single module (Table 1). Furthermore, temporal consistency in module identity was also

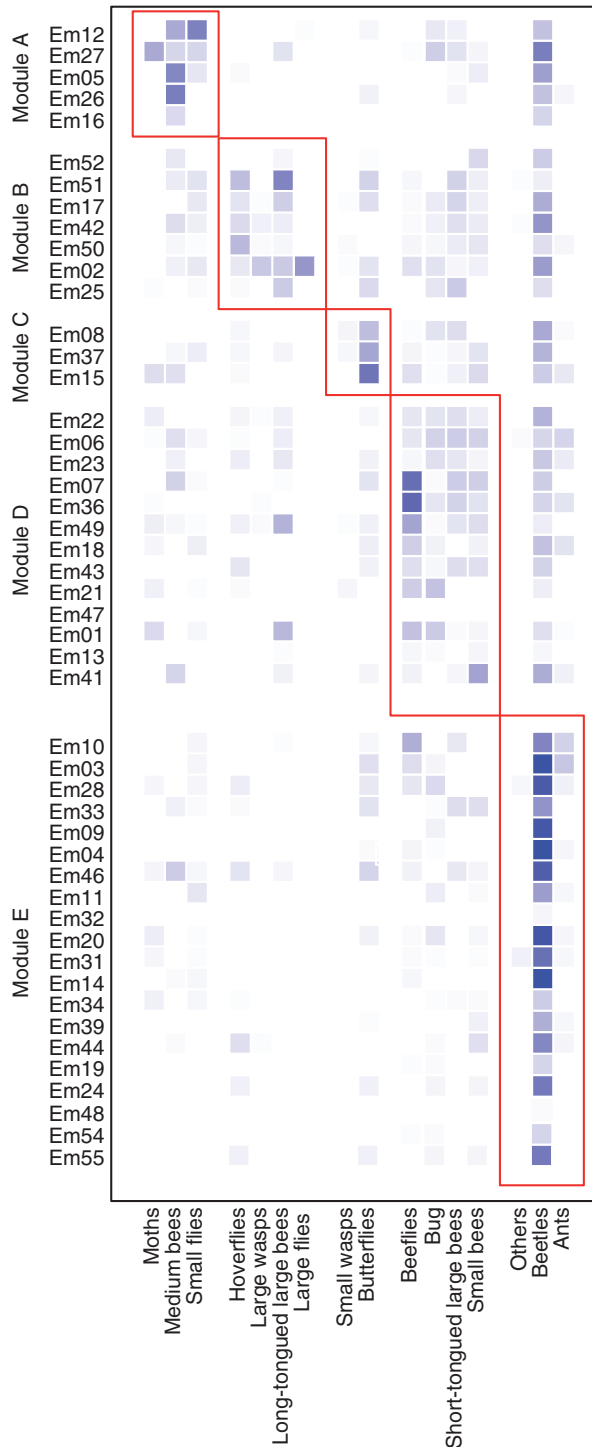


FIG. 2. Plot showing the average classification ($n = 50$ replicates) of the study populations into different pollinator modules, according to the analysis of bipartite modularity. The intensity of the colours indicates the relative abundance of each flower visitor's functional group per population.

high. Only five populations changed between modules, depending on the year. In fact, yearly modularity indices were as high as the overall index (2007, $M = 0.321$; 2008, $M = 0.261$; 2009, $M = 0.345$; 2010, $M = 0.334$).

TABLE 1. Frequency of each *Erysimum mediohispanicum* population belonging to each module when the modularity analysis is repeated 50 times

Population	Frequency of belonging to each module					Module ascription
	A	B	C	D	E	
Em01	0	0.12	0	0.88	0	D
Em02	0	0.96	0	0.04	0	B
Em03	0	0	0	0	1.00	E
Em04	0	0	0	0	1.00	E
Em05	1.00	0	0	0	0	A
Em06	0	0.14	0	0.86	0	D
Em07	0	0.02	0	0.98	0	D
Em08	0	0	1.00	0	0	C
Em09	0	0	0	0	1.00	E
Em10	0	0	0	0.36	0.64	E
Em11	0	0	0	0	1.00	E
Em12	1.00	0	0	0	0	A
Em13	0	0.02	0	0.98	0	D
Em14	0	0	0	0	1.00	E
Em15	0	0	1.00	0	0	C
Em16	1.00	0	0	0	0	A
Em17	0	1.00	0	0	0	B
Em18	0	0.02	0	0.98	0	D
Em19	0	0	0	0	1.00	E
Em20	0	0	0	0	1.00	E
Em21	0	0.12	0	0.88	0	D
Em22	0	0.18	0	0.82	0	D
Em23	0	0.18	0	0.82	0	D
Em24	0	0	0	0	1.00	E
Em25	0	1.00	0	0	0	B
Em26	1.00	0	0	0	0	A
Em27	0.60	0.26	0	0.14	0	A
Em28	0	0	0	0	1.00	E
Em31	0	0	0	0	1.00	E
Em32	0	0	0	0	1.00	E
Em33	0	0	0	0	1.00	E
Em34	0	0	0	0	1.00	E
Em36	0	0.02	0	0.98	0	D
Em37	0	0	1.00	0	0	C
Em39	0	0	0	0	1.00	E
Em41	0	0.04	0	0.96	0	D
Em42	0.38	0.62	0	0	0	B
Em43	0	0.08	0	0.92	0	D
Em44	0	0	0	0	1.00	E
Em46	0	0	0	0	1.00	E
Em47	0	0.02	0	0.98	0	D
Em48	0	0	0	0	1.00	E
Em49	0	0.08	0	0.92	0	D
Em50	0	1.00	0	0	0	B
Em51	0	1.00	0	0	0	B
Em52	0.28	0.44	0	0.28	0	B
Em54	0	0.02	0	0.16	0.82	E
Em55	0	0	0	0	1.00	E

The module to which each population was ascribed is also shown.

Spatial structure and phylogenetic signal of pollination niches

Neither the region ($\chi^2 = 7.47$, $P = 0.113$) nor the area ($\chi^2 = 10.32$, $P = 0.243$; generalized linear model) affected module identity. Populations belonging to all modules except modules A and C were found in both geographic regions occupied by the species and in the four study areas (Fig. 1). There was no phylogenetic signal in pollinator functional groups ($r = -0.04$, $P = 0.98$; partial Mantel test). As shown in Fig. 3, there were populations belonging to most of the modules in all of the

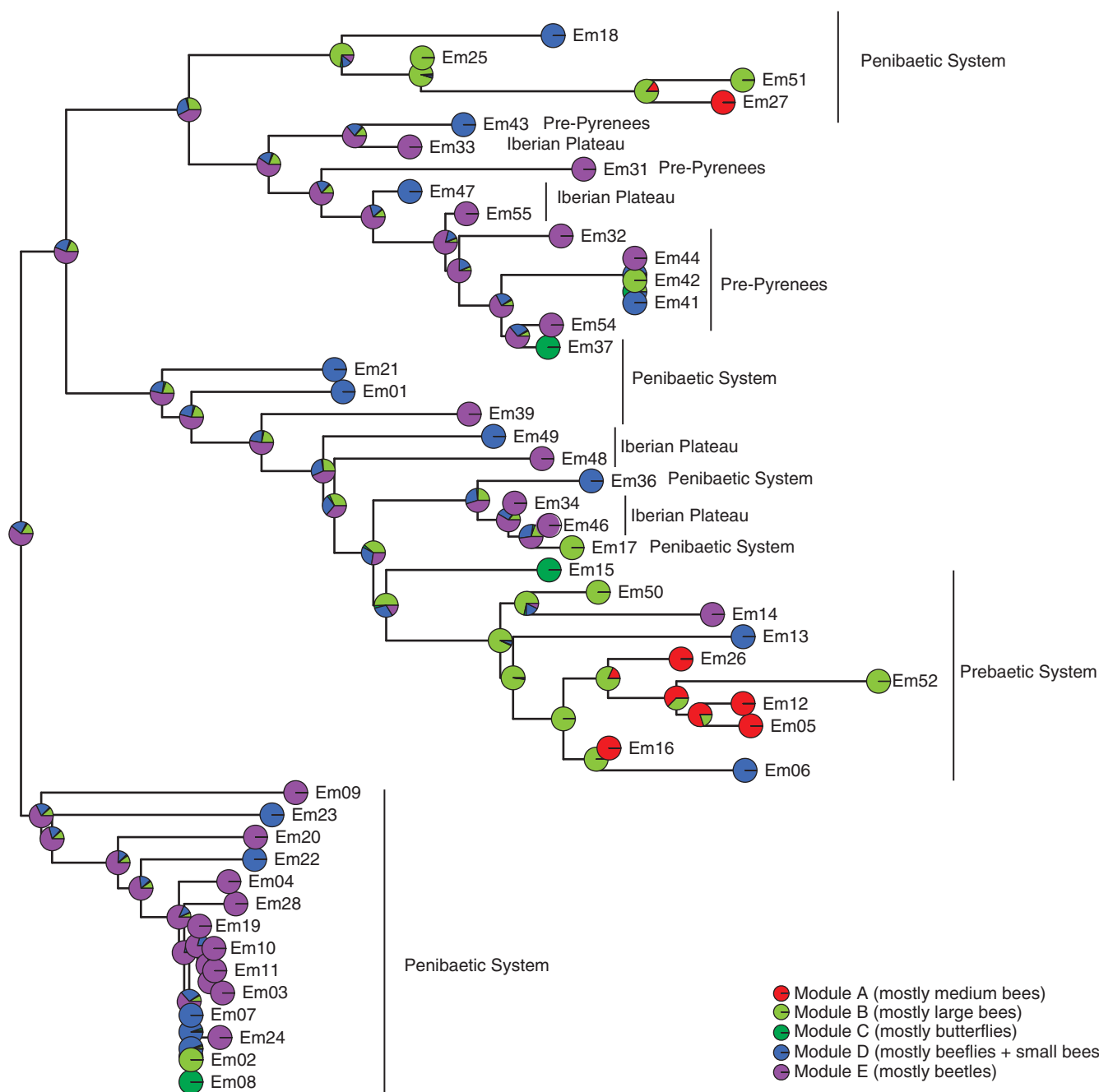


FIG. 3. Ancestral reconstruction of *Erysimum mediohispanicum* pollination niches according to the NJ tree describing the population genealogical relationship. The module identity of the terminal nodes (present-day populations) was considered as a fixed state (see Fig. 2). Different colours in the pie graphs of the internal nodes represent their distribution of posterior probability of belonging to each of the five modules.

three main *E. mediohispanicum* lineages (one including populations from the Penibaetic System, mostly Sierra Nevada, another including populations from the Penibaetic System and Pre-Pyrenees, and a third including a few populations from the Penibaetic System and all the populations from the Iberian Plateau and Prebaetic System; Supplementary Data Table S1), although populations from modules D and E appeared to be more frequent in the lineage from the Penibaetic System (Fig. 3). Populations from module A also appeared to be restricted mostly to the Prebaetic System (Fig. 3).

Ancestral reconstruction of pollination niches and transition rates

When the module identity of each *E. mediohispanicum* population was considered to be a fixed state, the reconstruction method suggested that the ancestral populations were visited mostly by beetles and ants (module E) or by beeflies and small bees (module D), since the posterior probabilities of the most ancestral node of belonging to these two modules ranged between 0.57 and 0.71 and between 0.17 and 0.35, respectively (Table 2). Module B (long-tongued large bees) was also reconstructed as the potential ancestral module in some chains (Table 2).

TABLE 2. Probability of the ancestral node in the *E. mediohispanicum* NJ tree belonging to each of the modules when (A) considering the module ascription of the populations as a fixed state; (B) including the uncertainty in module identity originated by repeating the modularity analysis 50 times; and (C) including the uncertainty in module identity originated by performing the modularity analysis separately each year

Population	Probability of belonging to each module				
	A	B	C	D	E
(A) Module identity of populations as a fixed state					
Chain 1	0	0.11	0.01	0.17	0.71
Chain 2	0	0.08	0.04	0.30	0.57
Chain 3	0.01	0.04	0.02	0.25	0.68
Chain 4	0	0.09	0.01	0.19	0.71
(B) Uncertainty in the module identity of the populations					
Chain 1	0	0.09	0.04	0.18	0.69
Chain 2	0	0.12	0.04	0.15	0.69
Chain 3	0	0.08	0.04	0.15	0.73
Chain 4	0	0.10	0.05	0.20	0.65
(C) Temporal uncertainty in module identity of the populations					
Chain 1	0	0.13	0.02	0.18	0.67
Chain 2	0	0.15	0.06	0.29	0.50
Chain 3	0	0.15	0.06	0.31	0.48
Chain 4	0	0.20	0.08	0.31	0.42

Populations visited mostly by butterflies (module C) and by medium-sized bees (module A) seem to have appeared later during the evolutionary history of *E. mediohispanicum*. This is especially true for module A. Moreover, it seems that on both occasions populations belonging to module A have appeared, presumably having evolved from populations belonging to module B (Fig. 3). Most other modules have appeared many times during the evolutionary history of *E. mediohispanicum* (Fig. 3; see Supplementary Data Table S3).

The ancestral reconstruction of modules was very similar when both computational and temporal uncertainty were included in the module identity of *E. mediohispanicum* populations (Table 2). The most ancestral *E. mediohispanicum* populations were again reconstructed as belonging to the modules visited by beetles/ants or by beesflies/small bees (Table 1), although the probability of belonging to module B was also considerable (Table 2; see Supplementary Data Tables S4 and S5). Again, modules A and C seemed to have appeared later in the evolutionary history of *E. mediohispanicum* (Supplementary Data Figs S2 and S3).

Correlated evolution of pollination niche and phenotypic traits

The number of flowers, corolla diameter, and the RW1 and RW2 shape components were correlated with the identity of the module when controlling both for across-population genetic relationship and for spatial distance (Table 3). Populations from module B had plants with more flowers than the rest of the modules (Fig. 4). Moreover, populations from modules B, D and E had larger corollas than did plants from modules A and C (Fig. 4). Finally, populations from module A had plants with open petals (positive RW1 shape component), whereas populations from modules B and D had plants with zygomorphic flowers (positive RW2 shape component; Fig. 4).

TABLE 3. Outcome of the MCMC generalized linear mixed models controlling for genetic relatedness and spatial location of the populations

Traits	Posterior mean	95 % confidence interval	MCMC P-value
Flower number	65.9957	42.49 to 87.81	0.0001
Stalk height	-22.6974	-51.52 to 10.14	0.166
Corolla diameter	17.3994	0.42 to 32.76	0.049
Corolla tube length	11.6836	-7.92 to 29.41	0.258
RW1	-322.2784	-593.25 to -97.01	0.0001
RW2	-743.8254	-1087.48 to -361.84	0.0001
RW3	328.452	-237.03 to 1015.57	0.333
RW4	-136.9831	-677.01 to 334.14	0.716

Deviance information criterion (DIC) = 114.30.

DISCUSSION

The existence of different pollination niches in a generalist plant

The flower visitors of *E. mediohispanicum* vary between populations in both diversity and identity (Gómez et al., 2007, 2009a). Between-population variation in pollinator assemblages is common in most generalist plants (Aigner, 2005; Moeller, 2005; Price et al., 2005; Burkle and Alarcon, 2011; Rader et al., 2012; and references therein). However, our network analysis indicates that the spatial variation in *E. mediohispanicum* pollinators was not random but rather was organized across populations. The 48 *E. mediohispanicum* populations surveyed in this study, despite being generalists, can be assembled into five modules according to their quantitative similarity in the type of insects visiting their flowers. It is possible to envisage a multidimensional space determined by orthogonal axes defined by changes in the abundance of each flower visitor type (Pauw, 2013). Each module would occupy a specific region within this niche space. Seen from this perspective, and if it is taken into account that flower visitors are resources for plants (Pauw, 2013), each module can be considered a different Eltonian pollination niche of *E. mediohispanicum* (Soberon, 2007). Our study suggests the occurrence of intraspecific, between-population variation in pollination niches in *E. mediohispanicum*. As a consequence of belonging to the same species, the boundaries between niches become fuzzy, and the degree of overlap between these five pollination niches is high (Wiens and Graham, 2005). Indeed, most populations were visited by functional insect groups associated with more than one pollination niche (Fig. 2; see also Supplementary Data Table S2). Furthermore, most flower visitors – such as beesflies, small bees, bugs and, above all, beetles – appeared in almost all populations, with only some functional groups (such as moths, butterflies, hoverflies or large wasps) restricted to a few populations (Fig. 2). However, despite this background of between-population variation in pollinators, we found that the ascription of each population to a given pollination niche was very robust.

It is tempting to consider populations from different pollination niches as belonging to different pollination syndromes. In fact, network modularity can reveal pollination syndromes in some systems (Dick et al., 2002; Danielli-Silva et al., 2012). However, we are not advocating the idea of generalist pollination

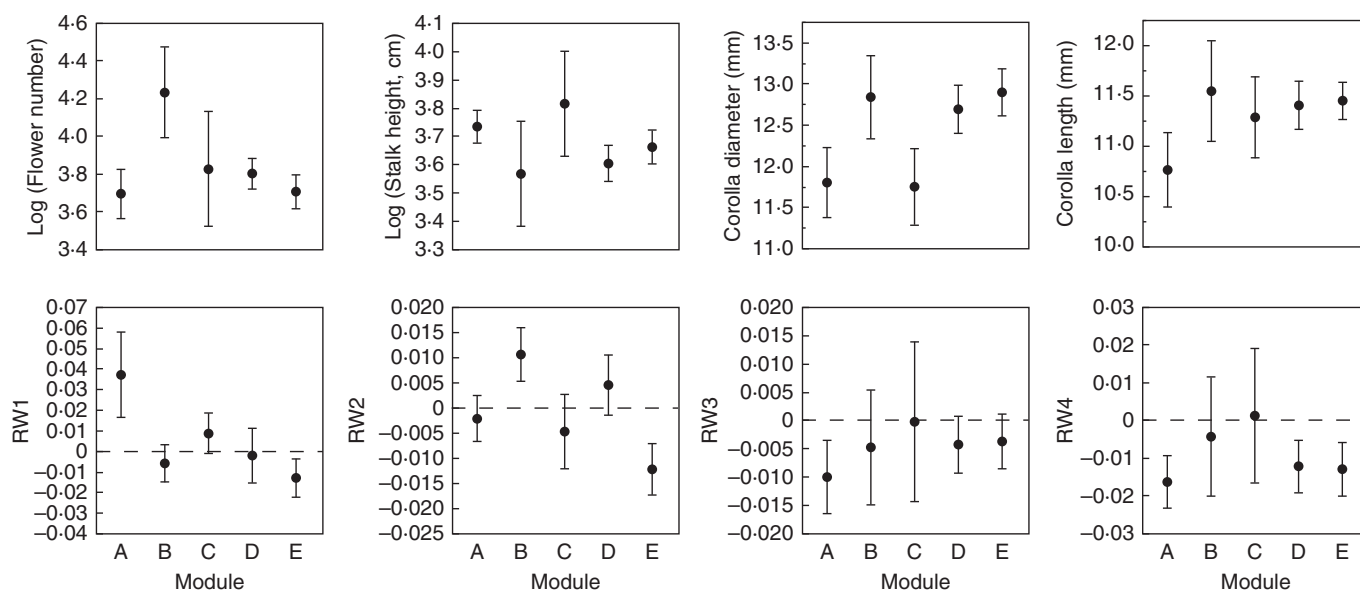


FIG. 4. Differences among pollination niches in the floral traits of *Erysimum mediohispanicum*. See Table 3 for statistical results using MCMC generalized linear mixed models. RW = relative warp (see Methods).

syndromes. Unlike true syndromes, the pollination niches detected in this study were not caused by the turnover of major functional groups of pollinators but by changes in the relative abundance of their floral visitors. Also, all the populations studied remained generalist. We did not detect any evolution towards increased specialization.

The floral visitors of *E. mediohispanicum* differed in their efficiency as pollinators. For example, although beetles are the main pollinators of *E. mediohispanicum* in some populations (Gómez, 2003) and ants can also act as pollinators of several coexisting crucifers (Gómez and Zamora, 1999), these insects are low-efficiency pollinators and behave more frequently as nectar thieves, pollen feeders and florivores than as legitimate pollinators (Leavitt and Robertson, 2006; Rico-Gray and Oliveira, 2007; Cardel and Koptur, 2010). The importance of these insects as pollinators strongly depends on how frequently they visit flowers rather than their per-visit effectiveness (Gómez and Zamora, 1999; Gómez et al., 2010). Other insects showing very low efficiency as pollinators of *Erysimum* spp. and other Brassicaceae are small and large flies, moths, bugs and small wasps (Gómez et al., 2009a, 2010; Lay et al., 2011; Fernández et al., 2012; Rader et al., 2012). Most of these insects frequently act as nectar thieves, entering the flowers from the bottom part of the corolla tube without making contact with the sexual organs (Gómez et al., 2009a). Short-tongued small, medium-sized and large bees, hoverflies and bee flies, unlike all the above-mentioned insects, can be efficient pollinators of *E. mediohispanicum* (Gómez et al., 2009a, 2010), other *Erysimum* species (Lay et al., 2011; Fernández et al., 2012) and many other crucifers (Rush et al., 1995; Cruden et al., 1996; Sahli and Conner, 2007; Rader et al., 2009, 2013; Robertson and Leavitt, 2011; Jauker et al., 2012). Although they sometimes act as pollen and nectar thieves, they act as legitimate pollinators most of the time. However, the most efficient pollinators of *Erysimum*, including *E. mediohispanicum*, seem to be long-tongued large bees. These insects move very fast,

visiting many flowers per time unit but few flowers within the same individual plant, and deliver many pollen grains per visit (Ollerton et al., 2007; Gómez et al., 2010; Lay et al., 2011). In fact, the *E. mediohispanicum* pollinators can be ranked in descending order according to their per-visit effectiveness (quantified as seed production), from long-tongued large bees, to short-tongued large bees, large wasps, medium-sized bees, bee flies, small bees, beetles, hoverflies, butterflies and flies (J. Valverde, Universidad de Granada, Spain, unpubl. data). Consequently, spatial variation in these types of insect causes geographical variations in the pollen limitation intensity and reproductive output, not only in *E. mediohispanicum* (Gómez et al., 2009a, 2010) but also in other congeneric species (*E. popovii*, Fernández et al., 2012; *E. capitatum*, Lay et al., 2011). Accordingly, belonging to a given pollination niche would have important consequences for the reproductive output of the plants.

The evolutionary history of *E. mediohispanicum* pollination niches

Variation in pollinator niches was not spatially structured but instead occurred at multiple geographic scales: between nearby populations, between areas within a geographic region and between regions. This finding suggests that, rather than varying as a cline (Anderson et al., 2009; Santos-Gally et al., 2013), the pollinator assemblages of *E. mediohispanicum* form a mosaic (Grant and Grant, 1965). The presence of a geographic mosaic in pollination environments seems to be frequent in generalist plants (Grant and Grant, 1965; Dilley et al., 2000). Furthermore, there was no phylogenetic signal in the evolution of the pollination niches. In fact, most pollination niches were scattered through both the geographic range and evolutionary history of *E. mediohispanicum* populations. This was especially true of the three scarce pollination niches (A, B and C), since populations from any of the three lineages were assigned to these niches. This pattern was also found, however, in the most common pollination niches (D and E). A few populations from

the Prebaetic System belonged to these two modules, although most of the remaining populations in that area (and clade) were ascribed to modules A and B. Altogether, these outcomes suggest that the evolution of the *E. mediohispanicum* pollination niches is labile. Many evolutionary and ecological factors may cause low phylogenetic conservatism and high lability in niche evolution (Losos, 2008; Cooper et al., 2010; Crisp and Cook, 2012), such as the occurrence of non-stabilizing selection – whether convergent or divergent – and local adaptation, low linkage of co-adapted traits, high genetic variation in niche-associated traits and a strong opportunity for the dispersal of species and populations (Crisp and Cook, 2012). More information is needed to disentangle these potential causes, although previous studies have indicated that divergent selection and local adaptation to pollinators is indeed occurring in some parts of the distribution area of *E. mediohispanicum* (Gómez et al., 2009a, b).

It appears that the ancestral *E. mediohispanicum* populations belonged to pollination niches E and, to a lesser extent, D. Thus, beetles, ants, bees, and short-tongued large and small bees were the most likely flower visitors of ancestral populations. It bears noting that the Penibaetic System lineage, the most ancient *E. mediohispanicum* lineage according to a phylogeographic study (Muñoz-Pajares, 2013), is composed almost entirely of populations belonging to these ancestral niches. Many other populations from other lineages and geographic areas also belong to these ancestral niches. Furthermore, we have even found several changes to these niches over the course of the evolutionary history of *E. mediohispanicum*, despite their being associated with pollinators with low or intermediate efficiency. Our analysis also suggests that there have been numerous shifts to other pollination niches during the evolution of *E. mediohispanicum*. In particular, the pollination niches B and C seem to have appeared several times, whereas niche C, being presumably of low efficiency, appeared three times. Altogether, the frequency of populations belonging to high-efficiency niches seems not to have increased during the evolution of *E. mediohispanicum*.

Pollinator-mediated floral divergence in *Erysimum mediohispanicum*

Phylogenetic changes in pollinators have been explored for many systems (e.g. *Disa*, Johnson et al., 1998; *Schizanthus*, Peres et al., 2006; *Aquilegia*, Whittall and Hodges, 2007; *Bastida* et al., 2010; *Iochroma*, Smith et al., 2008; *Ruellia*, Tripp and Manos, 2008). These studies have shown that shifts between pollination systems normally entail concomitant changes in floral phenotype. However, most of these studies have focused on the transitions between pollination syndromes found in various specialized plant species with divergent phenotypes. In contrast, it is widely assumed that pollinators play a minor role in the evolution and diversification of generalist plant clades because they do not provide strong reproductive isolation between diverging plant populations or pollination races in sympatry (Waser, 1998, 2001; Waser and Campbell, 2004; Kay and Sargent, 2009). Our study, however, suggests that a similar distinctive evolution may also occur during the evolutionary history of a single generalist plant species. In fact, we found a significant association between pollination niches and some *E. mediohispanicum* phenotypic traits. Plants from the

pollination niches D and E were very similar, having few and large flowers (Fig. 4), whereas plants from pollination niches A and C had smaller flowers, and plants from pollination niche B had many and large, high-rewarding flowers with highly zygomorphic (positive RW2) corollas. It is difficult to know whether these changes in floral size and shape are adaptations to local pollinators and can be considered pollination ecotypes, or on the contrary are due to other extrinsic factors. Biotic and abiotic agents, such as herbivores or altitude, also trigger phenotypic changes in *E. mediohispanicum* (Gómez, 2003). We presume, however, that pollinators may be at least partially responsible for some of the observed phenotypic differences. In fact, corolla size and the RW2 shape component are associated in this plant species not only with attractiveness but also with reward, i.e. pollen and nectar production (Gómez et al., 2008b). Furthermore, in a sub-set of eight populations from Sierra Nevada (south-east Spain) studied during 2005 and 2006, we found a geographic mosaic of selection produced by the quantitative spatial variation in pollinators with opposing preferences for *E. mediohispanicum* phenotypes (Gómez et al., 2008a, b, 2009a). Due to this geographic mosaic of selection, we detected divergent selection across some populations (Gómez et al., 2009a). A reciprocal translocation experiment even showed that this spatial variation in pollinator fauna causes the adaptation of plants to local pollinators in some localities (Gómez et al., 2009b). That is, there is experimental verification that changes in some *E. mediohispanicum* floral traits may be the consequence of local adaptation to contrasting pollinator faunas. For these reasons, we conjecture that geographic variation in pollination niches has contributed to the observed floral divergence in *E. mediohispanicum*.

It is important to establish how this floral variation has occurred. We presume that the process mediating this floral divergence has been similar to adaptive wandering (Thomson and Wilson, 2008). Assuming that our ancestral reconstruction analysis is correct, the *E. mediohispanicum* ancestral flowers (most probably belonging to pollination niche E) were large and symmetric. When *E. mediohispanicum* colonized new areas in the Iberian Peninsula, it found new, slightly different, although still generalist, pollination environments. Consequently, the plant faced contrasting pollinator-mediated selective scenarios. In some scarce localities where the pollinator fauna exerted strong and consistent selective pressures and where no other factors arrested this selection (selective hotspots; Gómez et al., 2009a), plants locally adapted to these newly found pollination niches (Gómez et al. 2009b), and generalist pollination ecotypes emerged. However, because the boundaries between generalist pollination niches are not sharp, the pollination ecotypes would differ, not abruptly, but gradually in a subtle way. The evolution of plants with many zygomorphic flowers in populations from pollination niche B would be an example of this type of pollination ecotype in *E. mediohispanicum*. In a previous experiment, we found that *E. mediohispanicum* was locally adapted to large bees through changes in, among other traits, the number of flowers and corolla zygomorphy (Gómez et al., 2009b). In addition, we presume that pollination ecotypes have not arisen universally across the plant's distribution area. In most localities, where pollination niches were sub-optimal, of low efficiency and unable to exert strong selection (selective coldspots; Gómez et al., 2009a), plants have not adapted locally to their pollinator fauna. The outcome is a geographic mosaic with a few populations

locally adapted to their pollination environment, where pollinator-driven floral divergence is occurring, coexisting with many populations where this local adaptation has failed and where the plant phenotype is not primarily shaped by pollinators. It is vital to gather further intra- and interspecific evidence to ascertain whether the scenario found in this study is idiosyncratic of some species or is a general pattern for pollinator-mediated floral evolution in generalist plants.

SUPPLEMENTARY DATA

Supplementary data are available online at www.aob.oxfordjournals.org and consist of the following. Methods: determining the phylogenetic relationship between *E. mediohispanicum* populations. Figure S1: summary of the geometric morphometric analysis. Figure S2: ancestral reconstruction of pollination niches including computational uncertainty of belonging to each module according to 50 replicates. Figure S3: ancestral reconstruction of pollination niches including temporal uncertainty of belonging to each module according to 50 replicates. Table S1: details of study populations. Table S2: frequency of each flower visitor functional group in each *Erysimum mediohispanicum* population. Table S3: posterior probability of internal nodes calculated using the ‘ace’ command in the R package phytool for both MCMC chains. Table S4: posterior probability of internal nodes calculated using the ‘ace’ command in the R package phytool for both MCMC chains when including a prior probability of each character state (= module) for each tip (= *E. mediohispanicum* population) resulting from running 50 networks. Table S5: posterior probability of internal nodes calculated using the ‘ace’ command in the R package phytool for both MCMC chains when including a prior probability of each character state (= module) for each tip (= *E. mediohispanicum* population) resulting from running a different network each year (2007–2010).

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LITERATURE CITED

- Abe T.** 2006. Threatened pollination systems in native flora of the Ogasawara (Bonin) Islands. *Annals of Botany* **98**: 317–334.
- Adams DC, Rohlf FJ, Slice DE.** 2004. Geometric morphometrics: ten years of progress following the ‘revolution’. *Italian Journal of Zoology* **71**: 5–16.
- Aigner PA.** 2005. Variation in pollination performance gradients in a *Dudleya* species complex: can generalization promote floral divergence? *Functional Ecology* **19**: 681–689.
- Anderson B, Alexandersson R., Johnson SD.** 2009. Evolution and coexistence of pollination ecotypes in an African *Gladiolus* (Iridaceae). *Evolution* **64**: 960–972.
- Armbruster WS.** 1993. Evolution of plant pollination systems: hypotheses and tests with the neotropical vine *Dalechampia*. *Evolution* **47**: 1480–505.
- Armbruster WS, Muchhala N.** 2009. Associations between floral specialization and species diversity: cause, effect, or correlation? *Evolutionary Ecology* **23**: 159–179.
- Armbruster WS, Edwards ME, Debevec EM.** 1994. Character displacement generates assemblage structure of Western Australian triggerplants (*Stylidium*). *Ecology* **75**: 315–29.
- Bastida JM, Alcántara JM, Rey PJ, Vargas P, Herrera CM.** 2010. Extended phylogeny of *Aquilegia*: the biogeographical and ecological patterns of two simultaneous but contrasting radiations. *Plant Systematics and Evolution* **284**: 171–185.
- Bates D.** 2005. Fitting linear mixing model in R using the lme4 package. *R News* **5**: 27–30.
- Burkle LA, Alarcón R.** 2011. The future of plant–pollinator diversity: understanding interaction networks across time, space, and global change. *American Journal of Botany* **98**: 528–538.
- Cardel Y, Koptur S.** 2010. Effects of florivory on the pollination of flowers: an experimental field study with a perennial plant. *International Journal of Plant Sciences* **171**: 283–292.
- Castellanos C, Wilson P, Thompson JD.** 2003. Pollen transfer by hummingbirds and bumblebees, and the divergence of pollination modes in *Penstemon*. *Evolution* **57**: 2742–2752.
- Cooley AM, Carvallo G, Willis JH.** 2008. Is floral diversification associated with pollinator divergence? Flower shape, flower color and pollinator preference in Chilean *Mimulus*. *Annals of Botany* **101**: 641–650.
- Cooper N, Jetz W, Freckleton RP.** 2010. Phylogenetic comparative approaches for studying niche conservatism. *Journal of Evolutionary Biology* **23**: 2529–2539.
- Crisp MD, Cook LG.** 2012. Phylogenetic niche conservatism: what are the underlying evolutionary and ecological causes? *New Phytologist* **196**: 681–694.
- Cruden RW, McClain AM, Shrivastava GP.** 1996. Pollination biology and breeding system of *Alliaria petiolata* (Brassicaceae). *Bulletin of the Torrey Botanical Club* **123**: 273–280.
- Danieli-Silva A, de Souza JMT, Donatti AJ, et al.** 2012. Do pollination syndromes cause modularity and predict interactions in a pollination network in a tropical high-altitude grassland? *Oikos* **121**: 35–43.
- Darwin CR.** 1862. *On the various contrivances by which British and foreign orchids are fertilised by insects*. London: John Murray.
- Dick CW, Etchelecu G, Austerlitz F.** 2003. Pollen dispersal of tropical trees (*Dinizia excelsa*: Fabaceae) by native insects and African honeybees in pristine and fragmented Amazonian rainforest. *Molecular Ecology* **12**: 753–764.
- Dilley JD, Wilson P, Mesler MR.** 2000. The radiation of *Calochortus*: generalist flowers moving through a mosaic of potential pollinators. *Oikos* **89**: 209–222.
- Dormann CF, Gruber B.** 2012. *Bipartite. Visualising bipartite networks and calculating some (ecological) indices*. Version 1-18. <http://cran.r-project.org/web/packages/bipartite> (6 February 2013).
- Dormann CF, Strauss R.** 2013. *Detecting modules in quantitative bipartite networks: the QuaBiMo algorithm*. ArXiv, 1304.3218v1.
- Faegri K, van der Pijl L.** 1979. *The principles of pollination ecology*. Oxford: Pergamon Press.
- Felsenstein J.** 2012. A comparative method for both discrete and continuous characters using the threshold model. *American Naturalist* **179**: 145–156.
- Fenster CB, Armbruster WS, Wilson P, Dudash MR, Thomson JD.** 2004. Pollination syndromes and floral specialization. *Annual Review of Ecology and Systematics* **35**: 375–403.

- Fernández JD, Bosch J, Nieto-Ariza B, Gómez JM. 2012. Pollen limitation in a narrow endemic plant: geographical variation and driving factors. *Oecologia* **170**: 421–431.
- Galen C, Cuba J. 2001. Down the tube: pollinators, predators, and the evolution of flower shape in the alpine skypilot, *Polemonium viscosum*. *Evolution* **55**: 1963–1971.
- Geib JC, Galen C. 2012. Tracing impacts of partner abundance in facultative pollination mutualism: from individuals to populations. *Ecology* **93**: 1581–1592.
- Gómez JM. 2003. Herbivory reduces the strength of pollinator-mediated selection in the Mediterranean herb *Erysimum mediohispanicum*: consequences for plant specialization. *American Naturalist* **162**: 242–256.
- Gómez JM, Perfectti F. 2010. Evolution of complex traits: the case of *Erysimum* corolla shape. *International Journal of Plant Sciences* **171**: 987–998.
- Gómez JM, Zamora R. 1999. Generalization in the interaction between *Hormathophylla spinosa* (Cruciferae) and its pollinators. *Ecology* **80**: 796–805.
- Gómez JM, Bosch J, Perfectti P, Fernández JD, Abdelaziz M. 2007. Pollinator diversity affects plant reproduction and recruitment: the tradeoffs of generalization. *Oecologia* **153**: 597–605.
- Gómez JM, Bosch J, Perfectti F, Fernández JD, Abdelaziz M, Camacho JPM. 2008a. Spatial variation in selection on corolla shape in a generalist plant is promoted by the preference patterns of its local pollinators. *Proceedings of the Royal Society B: Biological Sciences* **275**: 2241–2249.
- Gómez JM, Bosch J, Perfectti F, Fernández JD, Abdelaziz M, Camacho JPM. 2008b. Association between floral traits and reward in *Erysimum mediohispanicum* (Brassicaceae). *Annals of Botany* **101**: 1413–1420.
- Gómez JM, Perfectti F, Bosch J, Camacho JPM. 2009a. A geographic selection mosaic in a generalized plant–pollinator–herbivore system. *Ecological Monographs* **79**: 245–264.
- Gómez JM, Abdelaziz M, Camacho JPM, Muñoz-Pajares J, Perfectti F. 2009b. Local adaptation and maladaptation to pollinators in a generalist geographic mosaic. *Ecology Letters* **12**: 672–682.
- Gómez JM, Abdelaziz M, Lorite J, Muñoz-Pajares AJ, Perfectti F. 2010. Changes in pollinator fauna cause spatial variation in pollen limitation. *Journal of Ecology* **98**: 1243–1252.
- Grant V, Grant KA. 1965. *Flower pollination in the phlox family*. New York: Columbia University Press.
- Guimerà R, Amaral LA. 2005. Functional cartography of complex metabolic networks. *Nature* **433**: 895–900.
- Guimerà R, Sales-Pardo M, Amaral LA. 2007. Module identification in bipartite and directed networks. *Physical Review E* **76**: 036102.
- Hadfield JD. 2013. Package ‘MCMCglmm’. Version 2.12.7. <http://cran.r-project.org/web/packages/MCMCglmm> (15 February 2013).
- Hadfield JD, Nakagawa S. 2010. General quantitative genetic methods for comparative biology: phylogenies, taxonomies and multi-trait models for continuous and categorical characters. *Journal of Evolutionary Biology* **23**: 494–508.
- Harder LD, Johnson SD. 2009. Darwin’s beautiful contrivances: evolutionary and functional evidence for floral adaptation. *New Phytologist* **183**: 530–545.
- Hardy OJ, Pavoine S. 2012. Assessing phylogenetic signal with measurement error: a comparison of Mantel tests, Blomberg *et al.*’s *K*, and phylogenetic distograms. *Evolution* **66**: 2614–2621.
- Herrera CM. 1996. Floral traits and adaptation to insect pollinators; a devil’s advocate approach. In: Lloyd DG, Barrett SCH. eds. *Floral biology*. New York: Chapman & Hall, 65–87.
- Honnay O, Jacquemyn H, van Looy K, Vandepitte K, Breyne P. 2009. Temporal and spatial genetic variation in a metapopulation of the annual *Erysimum cheiranthoides* on stony river banks. *Journal of Ecology* **97**: 131–141.
- Jauker F, Bondarenko B, Becker HC, Steffan-Dewente I. 2012. Pollination efficiency of wild bees and hoverflies provided to oilseed rape. *Agricultural and Forest Entomology* **14**: 81–87.
- Johnson SD. 2006. Pollinator-driven speciation in plants. In: Harder LD, Barrett SCH. eds. *Ecology and evolution of flowers*. Oxford: Oxford University Press, 295–310.
- Johnson SD. 2010. The pollination niche and its role in the diversification and maintenance of the southern African flora. *Philosophical Transactions of the Royal Society B: Biological Sciences* **365**: 499–516.
- Johnson SD, Linder HP, Steiner KE. 1998. Phylogeny and radiation of pollination systems in *Disa* (Orchidaceae). *American Journal of Botany* **85**: 402–411.
- Kay KM, Sargent RD. 2009. The role of animal pollination in plant speciation: integrating ecology, geography, and genetics. *Annual Review of Ecology, Evolution, and Systematics* **40**: 637–656.
- Lay CR, Linhart YB, Diggle PK. 2011. The good, the bad and the flexible: plant interactions with pollinators and herbivores over space and time are moderated by plant compensatory responses. *Annals of Botany* **108**: 749–63.
- Leavitt H, Robertson IC. 2006. Petal herbivory by chrysomelid beetles (*Phyllotreta* sp.) is detrimental to pollination and seed production in *Lepidium papilliferum* (Brassicaceae). *Ecological Entomology* **31**: 657–660.
- Losos JB. 2008. Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. *Ecology Letters* **11**: 995–1003.
- Mardulyn P. 2012. Trees and/or networks to display intraspecific DNA sequence variation? *Molecular Ecology* **21**: 3385–3390.
- Medel R, Valiente A, Botto-Mahan C, *et al.* 2007. The influence of insects and hummingbirds on the geographical variation of the flower phenotype in *Mimulus luteus*. *Ecography* **30**: 812–818.
- Moeller DA. 2005. Pollinator community structure and sources of spatial variation in plant–pollinator interactions in *Clarkia xantiana* ssp. *xantiana*. *Oecologia* **142**: 28–37.
- Muñoz-Pajares AJ. 2013. SIDIER: substitution and indel distances to infer evolutionary relationships. *Methods in Ecology and Evolution*, in press.
- Newman MEJ. 2004. Analysis of weighted networks. *Physical Review E* **70**: 056131.
- van der Niet T, Pirie MD, Shuttleworth A, Johnson SD, Midgley JJ. 2014. Do pollinator distributions underlie the evolution of pollination ecotypes in the Cape shrub *Erica plukenetii*? *Annals of Botany* **113**: doi:10.1093/aob/mct193.
- Nieto-Feliner G. 1993. *Erysimum* L. In: Castroviejo S. ed. *Flora iberica. Vol. IV. Cruciferae-Monotropaceae*. Madrid: Real Jardín Botánico, CSIC, 48–76.
- Nunn CL. 2011. *The comparative method in evolutionary anthropology and biology*. Chicago: University of Chicago Press.
- Ollerton J, Stott A, Allnutt E, Shove S, Taylor C, Lamborn E. 2007. Pollination niche overlap between a parasitic plant and its host. *Oecologia* **151**: 473–485.
- Paradis E, Claude J, Strimmer K. 2004. APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**: 289–290.
- Parrish JAD, Bazzaz FA. 1978. Pollination niche separation in a winter annual community. *Oecologia* **35**: 133–140.
- Parrish JAD, Bazzaz FA. 1979. Differences in pollination niche relationships in early and late successional plant communities. *Ecology* **60**: 597–610.
- Pavoine R, Ricotta C. 2013. Testing for phylogenetic signal in biological traits: the ubiquity of cross-product statistics. *Evolution* **67**: 828–840.
- Pauw A. 2013. Can pollination niches facilitate plant coexistence? *Trends in Ecology and Evolution* **28**: 30–37.
- Peres F, Arroyo MTK, Medel R, Hershkovitz MA. 2006. Ancestral reconstruction of flower morphology and pollination systems in *Schizanthus* (Solanaceae). *American Journal of Botany* **93**: 1029–1038.
- Perez-Barrales R, Arroyo J, Armbruster WS. 2007. Differences in pollinator faunas may generate geographic differences in floral morphology and integration in *Narcissus papyraceus* (Amaryllidaceae). *Oikos* **116**: 1904–1918.
- Price MV, Waser NM, Irwin RE, Campbell RD, Brody AK. 2005. Temporal and spatial variation in pollination of a montane herb: a seven-year study. *Ecology* **86**: 2106–2116.
- Rader R, Howlett B, Cunningham SA, *et al.* 2009. Alternative pollinator taxa are equally efficient, but not as effective as the honeybee in a mass flowering crop. *Journal of Applied Ecology* **46**: 1080–1087.
- Rader R, Howlett B, Cunningham SA, Westcott D, Edwards W. 2012. Spatial and temporal variation in pollinator effectiveness: do unmanaged insects provide consistent pollination services to mass flowering crops? *Journal of Applied Ecology* **49**: 126–134.
- Rader R, Edwards W, Westcott D, Cunningham SA, Howlett B. 2013. Diurnal effectiveness of pollination by bees and flies in agricultural *Brassica rapa*: implications for ecosystem resilience. *Basic and Applied Ecology* **14**: 20–27.
- Revell LJ. 2012. phytools: an R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution* **3**: 217–223.
- Rico-Gray V, Oliveira PS. 2007. *The ecology and evolution of ant–plant interactions*. Chicago: University of Chicago Press.
- Robertson IC, Leavitt H. 2011. Relative contributions to seed production by floral visitors of slickspot peppergrass, *Lepidium papilliferum* (Brassicaceae). *Arthropod-Plant Interactions* **5**: 379–389.

- Sahli H, Conner JK. 2007.** Visitation, effectiveness, and efficiency of 15 genera of visitors to wild radish, *Raphanus raphanistrum* (Brassicaceae). *American Journal of Botany* **94**: 203–209.
- Sahli HF, Conner JK. 2011.** Testing for conflicting and nonadditive selection: floral adaptation to multiple pollinators through male and female fitness. *Evolution* **65**: 1457–1473.
- Santos-Gally R, Pérez-Barrales R, Simón VI, Arroyo J. 2013.** The role of short-tongued insects in floral variation across the range of a style-dimorphic plant. *Annals of Botany* **111**: 317–328.
- Sargent RD, Otto SP. 2006.** The role of local species abundance in the evolution of pollinator attraction in flowering plants. *American Naturalist* **167**: 67–80.
- Smith SD, Ane C, Baum DA. 2008.** The role of pollinator shifts in the floral diversification of *Iochroma* (Solanaceae). *Evolution* **62**: 793–806.
- Soberón J. 2007.** Grinnellian and Eltonian niches and geographic distributions of species. *Ecology Letters* **10**: 1115–1123.
- Stone GN, Nee S, Felsenstein J. 2011.** Controlling for non-independence in comparative analysis of patterns across populations within species. *Philosophical Transactions of the Royal Society B: Biological Sciences* **366**: 1410–1424.
- Thomson JD, Wilson P. 2008.** Explaining evolutionary shifts between bee and hummingbird pollination: convergence, divergence, and directionality. *International Journal of Plant Sciences* **169**: 23–38.
- Tripp EA, Manos PS. 2008.** Is floral specialization an evolutionary dead-end? Pollination system evolution in *Ruellia* (Acanthaceae). *Evolution* **62**: 1712–1737.
- Waser NM. 1998.** Pollination, angiosperm speciation, and the nature of species boundaries. *Oikos* **82**: 198–201.
- Waser NM. 2001.** Pollinator behavior and plant speciation: looking beyond the ‘ethological isolation’ paradigm. In: Chittka L, Thomson JD. eds. *Cognitive ecology of pollination*. Cambridge, UK: Cambridge University Press, 318–335.
- Waser NM, Campbell DR. 2004.** Ecological speciation in flowering plants. In: Dieckmann U, Doebeli M, Metz JAJ, Tautz D. eds. *Adaptive speciation*. Cambridge: Cambridge University Press, 264–277.
- Whittall JB, Hodges SA. 2007.** Pollinator shifts drive increasingly long nectar spurs in columbine flowers. *Nature* **447**: 706–710.
- Wiens JJ, Graham CH. 2005.** Niche conservatism: integrating evolution ecology and conservation biology. *Annual Review of Ecology, Evolution, and Systematics* **36**: 519–539.
- Willmer P. 2011.** *Pollination and floral ecology*. Princeton, NJ: Princeton University Press.
- Wilson P, Castellanos MC, Hogue JN, Thomson JD, Armbruster WS. 2004.** A multivariate search for pollination syndromes among penstemons. *Oikos* **104**: 345–361.
- Zelditch ML, Swinderski DL, Sheets HD, Fink WL. 2004.** *Geometric morphometrics for biologists: a primer*. San Diego: Elsevier Academic.